Bioscience Discovery, 9(4):474-477, Oct - 2018

© RUT Printer and Publisher Print & Online, Open Access, Research Journal Available on http://jbsd.in ISSN: 2229-3469 (Print); ISSN: 2231-024X (Online) Research Article



Isolation and Characterization of antibiotic producers from Vajreshwari Hot Spring, Mumbai

Mukundraj G. Rathod, Sandhya C. Dhembre, Sandip V. Thorat & ¹Anupama P. Pathak*

Dept. of Biotechnology & Bioinformatics (UG & PG), Center for Advanced Studies, Yeshwant College of IT (BI & BT), Parbhani 431401. ¹School of Life Sciences (DST, FIST & UGC-SAP Sponsored) ¹Swami Ramanand Teerth Marathwada University, Nanded 431606. * anupama.micro@rediffmail.com

Article Info

Abstract

Received: 23-05-2018, Revised: 02-09-2018, Accepted: 19-09-2018

Keywords:

Bacillus aeolius, Bacillus thermoamylovorans, crowded plate technique, thermostable enzyme, Vajreshwari hot spring

INTRODUCTION

Vajreshwari town (19°29'12"N 73°1'33"E) is situated on bank of the river Tansa, in Bhiwandi city of Thane district, Maharashtra, India. There are approximately 21 hot springs within 5 km radius of Vajreshwari shrine. It is reported that, temperature of these hot springs ranges from 43 to 80°C. Many religious people and tourists visit to this place (Patil and Unnikrishnan, 2015). Antibiotics producers mostly belong to various genera of filamentous actinomycetes and Gram positive bacilli (Gebrevohannes et al., 2013, Denver et al., 2004). According to literature thermal springs are habitats of many actinomycetes and bacilli. Streptomyces Micromonospora sp., sp., Microbispora sp. and Planosporangium sp. are antibiotic producing actinomycetes previously reported from Thai hot spring sediment by Thawai et al., (2005). Chaudhary and Prabhu, (2016) have previously reported thermophilic actinomycetes from Vajreshwari hot springs. Very few species of

and characterized thoroughly. On the basis of morphological characters, microscopic features and biochemical profile, VHSI-1 and VHSI-2 were identified as *Bacillus aeolius* and *Bacillus thermoamylovorans* respectively. These isolates can be explored for detection of possibly new promising antibiotic as well as production of various industrially important thermostable enzymes.

In the era of assessment of thermophiles and their characteristics, we have

selected Vajreshwari hot spring, Mumbai. Vajreshwari town (19°29'12"N

73°1′33″E) is located in Thane district, Maharashtra, India. Water sample from this hot spring was screened for isolation of antibiotic producers by crowded

plate technique. Two thermophiles namely, VHSI-1 and VHSI-2 were isolated

Bacillus from hot springs are known to produce antibiotic. In this context, we attempted isolation and characterization of antibiotic producers from Vajreshwari hot spring, Maharashtra. Moreover, the isolates reported by us have also shown the ability to produce a wide array of enzymes to be applied in biotechnological industries and biomedical fields.

MATERIALS AND METHODS

Isolation, characterization and preliminary identification:

Water samples from one of the Vajeshwari hot springs were collected in Jan. 2018. Temperature and pH of water sample was recorded *in situ* by using digital thermometer and pH meter respectively. Crowed plate technique was used for isolation of antibiotic producer (Aneja, 2006; Kamoldeen, 2013). In this method, water samples from Vajeshwari hot spring ranging in volume 0.1 to 2.0 mL were inoculated on nutrient agar plates aseptically. All the plates were incubated in a bacteriological incubator (Kumar make, Mumbai) at 65°C for 18 h for further studies. Morphological, microscopic, biochemical and physiological characters of selected isolate were determined as per the standard methods which are already described in the references. following cited Preliminary identification of selected isolates was carried out by using Bergey's Manual of systematic bacteriology (Vos et al., 2009; Rathod and Pathak, 2014; Rathod and Pathak, 2014a; Rathod and Pathak, 2016; Rathod and Pathak, 2016a; Pathak and Rathod, 2015; Pathak and Rathod, 2015a; Pathak and Rathod, 2013; Pathak and Rathod, 2014; Pathak et al; 2014, Pathak et al., 2015; Pathak et al., 2015a; Pathak and Rathod, 2016; Pathak *et al.*, 2016; Pathak *et al.*, 2016a; Pathak *et al.*, 2016b; Rathod and Pathak, 2016b; Rathod and Pathak, 2016c; Rathod and Pathak, 2017; Pathak and Rathod, 2016a; Pathak and Rathod, 2017).

RESULTS AND DISCUSSION

Isolation of antibiotic producer:

Temperature and pH of vajreshwari hot spring was recorded 65°C and 7.89 respectively. After incubation, two microbial colonies were observed that showed zone of growth inhibition (Table 1). These colonies were designated as VHSI-1 and VHSI-2 and further subcultured on nutrient agar slants.

Volume of water	Total colony	Designation of Antibiotic	size of zone of growth
sample inoculated	count	producers	inhibition (mm) in crowed
(mL)			plate technique
0.1	37	-	-
0.5	189	VHSI-1	11
1.0	292	VHSI-2 and VHSI-3	10 and 09
1.5 and 2.0	Mat growth	-	_

Table 1: Size of zone of growth inhibition

Characterization and preliminary identification:

Colonies of VHSI-1 were circular and cream-colored. VHSI-1 was Gram stain variable and motile rod. Length and width of a cell was 2.0 and 0.5 µm respectively. VHSI-1 formed terminally located endospore on sporulation medium. VHSI-1 was catalase-negative and oxidase positive. Temperature range for growth of VHSI-1 was found 35 to 65°C, with an optimum at 55 °C. pH range for growth of VHSI-1 was found 7 to 9, with an optimum at pH 8. NaCl concentration range for growth of VHSI-1 was found 0.5 to 5%, with an optimum at 2% NaCl. Acid without gas is produced by VHSI-1 from a wide range of carbohydrates viz. glucose, mannitol, mannose, salicin, starch and xylose. Casein, gelatin and starch substrates were hydrolyzed by VHSI-1. On the basis of morphological characters, microscopic features and biochemical profile, VHSI-1was identified as Bacillus aeolius. This organism was previously reported from a shallow marine hydrothermal vent, Vulcano Island and Eolian Islands, Italy (Vos et al. 2009).

Colonies of VHSI-2 were white, lenticular, and 2 mm in diameter. VHSI-2 was Gram stain positive and weakly motile rods. Length and width of a cell of VHSI-2 was 3.0 and 0.5μ m respectively. Endospore formation was not observed. VHSI-2

showed optimum growth at 60°C and pH 7. VHSI-2 was catalase-positive and oxidase-negative. Starch was efficiently hydrolyzed by VHSI-2 (Figure 1). Acid without gas was produced by VHSI-2 in presence of glucose and starch. On the basis of morphological characters, microscopic features and biochemical profile, VHSI-2 was identified as *Bacillus thermoamylovorans*. This organism was previously reported from palm wine (Vos et al. 2009). Identification of VHSI-3 isolate is in progress.



Figure 1: Starch hydrolysis by *Bacillus thermoamylovorans* VHSI-2 isolate from Vajreshwari hot spring, Maharashtra.

Conclusions

In conclusion, two bacterial isolates from Vajreshwari hot spring designated as VHSI-1 and VHSI-2 were found antibiotic producers as screened by crowded plate technique. On the basis of morphological characters, microscopic features and biochemical profile, VHSI-1 and VHSI-2 were identified as *Bacillus aeolius* and *Bacillus thermoamylovorans* respectively. These isolates can be explored for detection of possibly new promising antibiotic. Moreover production of various thermozymes can also be achieved from these isolates.

ACKNOWLEDGEMENTS

Former three authors are thankful to Prof. Dr. Anupama P. Pathak, Head of Microbiology Faculty, S.L.S., S.R.T.M. University, Nanded for her excellent guidance, innovative ideas and thoughts and also thankful to Hon. Dr. Rafiq Shaikh, Chairman of M.E.C.H. & W. Society, Parbhani for providing infrastructure and necessary facilities.

REFERENCES

Aneja KR, 2005. Experiments in Microbiology Plant Pathology and Biotechnology. New Age International (P) Ltd., New Delhi India.

Chaudhary N and Prabhu S, 2016. Thermophilic Actinomycetes from Hot Water Spring Capable of Producing Enzymes of Industrial Importance. *Int. J. Research Studies in Biosci.* **4**(6):29-35.

Denyer SP, Hodges NA and Gorman SP, 2004. *Hugo and Russell's Pharmaceutical Microbiology*, 7th Edition, Blackwell Publishing company.

Gebreyohannes G, Moges F, Sahile S and Raja N, 2013. Isolation and characterization of potential antibiotic producing actinomycetes from water and sediments of Lake Tana, Ethiopia. *Asian Pac. J. Trop. Biomed.* **3**(6): 426-435.

Kamoldeen A, 2013. Soil Screening for Antibiotic - Producing Microorganisms. *Adv. Env. Biol.* **7**(1):7-11.

Pathak AP and Rathod MG, 2013. Production and characterization of alkaline protease by *Bacillus pasteurii*: a Lonar soda lake isolate. *Innov. Res. Chem.* 1(1):22-26.

Pathak AP and Rathod MG, 2014. Exploration of Unkeshwar hot springs in Maharashtra for thermostable amylase producer. *Res. Rev. Biosci.* **8**(7): 269-276.

Pathak AP and Rathod MG, 2015. A report on thermostable alkaline protease producing bacteria

from a terrestrial thermal spring. *Indian J. Mar. Sci.* **44**(7):1104-1111.

Pathak AP and Rathod MG, 2015a. Cultivable bacterial diversity of terrestrial thermal spring of Unkeshwar. *J. Biochem. Tech.* **5**(4):814-818.

Pathak AP and Rathod MG, 2016. Assessment of diverse thermostable alkaline lipase producers from Unkeshwar hot spring of Maharashtra, India. *Concept. Pure Appl. Sci.* **3**(1):1-9.

Pathak AP and Rathod MG, 2016a. Production and characterization of thermostable gelatinase from *Bacillus globisporus* isolated from Unkeshwar hot spring of Maharashtra. *Indian J. Mar. Sci.* 46(9):1883-1888.

Pathak AP and Rathod MG, 2017. Exploration of a hot spring for thermostable protease producers. *J. Microbiol. Biotechnol. Food Sci.* **7**(2):101-109.

Pathak AP, Devkatte SB and Rathod MG, 2016b. Production of amylase by *Penicillium* sp. by using solid state fermentation method and inexpensive agricultural residue. *International journal for research in biology and pharmacy research.* 2(4):23-33.

Pathak AP, Lodge N, Gavali JT and Rathod MG, 2015. Isolation and characterization of coldactive protease producer from ice factory samples. *Int. J. Adv. Pharm. Biol. Chem.* 4(4):751-754.

Pathak AP, Lohagave AG and Rathod MG, 2015a. Exploration of paper industry effluent for isolation of efficient starchy material degrader to promote bioremediation. *Int. J. Adv. Pharm. Biol. Chem.* 4(4) 729-736.

Pathak AP, Mahindrakar PR and Rathod MG, 2016a. Defatted seed meal of soybean —an inexpensive substrate for alkaline protease production from selected isolate. *International journal for research in agricultural research*. 2(4):37-48.

Pathak AP, Parware SU and Rathod MG, 2016. Isolation and identification of industrially important salt stable amylase producer. *International journal for research in biology and pharmacy research*, **2**(3):41-51.

Pathak AP, Rathod MG and Rampurkar VV, 2014. An eco-friendly approach for thermostable amylase production using *Bacillus firmus* APP6: a hot spring isolate. *Asiatic J. Biotech. Res.* **4**(4): 101-105.

Patil S and Unnikrishnan G, 2015. Hot water springs: Ancient and Modern Era. J. Global Biosci. 4(6): 2468-2472.

Rathod MG and Pathak AP, 2014. Wealth from Waste: Optimized alkaline protease production using agro-industrial residues by *Bacillus alcalophilus* LW8 and its biotechnological applications. *J. Taibah Univ. Sci.* **8**(4):307–314.

Rathod MG and Pathak AP, 2014a. Isolation and identification of alkaline protease producers from selected alkaline habitat. *Int. J. Innov. Biol. Res.* **3**(1):1-6.

Rathod MG and Pathak AP, 2016. Optimized production, characterization and application of alkaline proteases from taxonomically assessed microbial isolates from Lonar soda lake, India. *Biocatal. Agric. Biotechnol.* **7**: 164-173.

Rathod MG and Pathak AP, 2016a. Data on optimized production and characterization of alkaline proteases from newly isolated alkaliphiles from Lonar soda lake, India. *Data Brief.* **8**(3): 863-866.

Rathod MG and Pathak AP 2016b. Production, extraction and characterization of cold active and

salt stable alkaline protease from *Halomonas* sp. LAP520: Lonar soda lake isolate. *Int. J. Adv. Res. Rev.* **1**(5):123-127.

Rathod MG and Pathak AP, 2016c. Taxonomic assessment of alkali tolerant metallophiles from soil of M.I.D.C. Parbhani. *Int. J. Adv. Res. Rev*, 1(6):29-39.

Rathod MG and Pathak AP, 2017. Evaluation of the enzymatic profile of microbial isolates from Lonar Soda Lake, *Indian J. Mar. Sci.* **46**(1):116-124.

Thawai C, Tanasupawat S, Itoh T, Suwanborirux K, Suzuki KI and Kudo T, 2005. *Micromonospora eburnea* sp. nov., isolated from a Thai peat swamp forest. *Int. J. Syst. Evol. Microbiol.* 55:417-422.

Vos PD, Garrity GM, Jones D, Krieg NR, Ludwig W, Rainey FA, Schleifer KH, Whitman WB and Parte AC, 2009. Bergey's Manual of Systematic bacteriology. 2nd Edition, Vol 3. Springer Dordrecht Heidelberg London New York.

How to cite this article

Mukundraj G. Rathod, Sandhya C. Dhembre, Sandip V. Thorat & Anupama P. Pathak, 2018. Isolation and Characterization of antibiotic producers from Vajreshwari Hot Spring, Mumbai. *Bioscience Discovery*, 9(4):474-477.