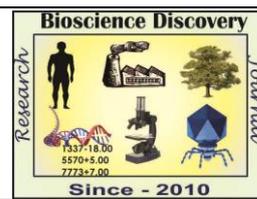


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Research Article



A comparative study on RNA: DNA Ratio in different tissues in two species of fresh water bivalve *Lamellidens* from lotic and lentic water

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Abstract

In the present study an attempt has been made to understand changes in RNA: DNA ratio in different tissues in relation to reproduction in freshwater water two species of bivalve *Lamellidens marginalis* from Godavari river and *Lamellidens corrianus* from Nandrabad pond in different seasons. Seasonal variation found in the ratio in both male and female tissues. In *Lamellidens marginalis* both males and females the ratio found gill > mantle > gonad > hepatopancreas in monsoon season. High ration found in hepatopancreas and gonads in winter in both sexes and in winter male gills and hepatopancreas have shown highest RNA: DNA ratio. In *Lamellidens corrianus* males tissues like gonad gill mental an hepatopancreas have higher RNA: DNA ratio in summer season than monsoon and winter and in females the ratio found gill, mantle, gonad and hepatopancreas have higher RNA:DNA ratio in monsoon season.

INTRODUCTION

Some of the aspects of the invasive process (number of colonization events, size of the founding population, pathway of the colonization, etc.) can be revealed by molecular biology studies, and as consequence, the use of molecular biology methods in studies of evolution and population genetics of invasive species has increased dramatically in the last years (Graputto *et al.*, 2005; Timmermans *et al.*, 2005). RNA: DNA ratio is now daysmost widely-used tool to determine biochemical and eco-physiological index of activity (growth, reproduction, secretion, etc.) of organisms under a given environmental condition (Lucas and Beninger, 1985). This index gives a measure of the synthetic capacity of the cell and usually correlates with nutritional status (Buckley *et al.*, 1999). The RNA: DNA ratio is based on the assumption that the amount of DNA, the primary carrier of genetic information, is stable under changing environmental

situations within the somatic cells of a species (Bulow, 1987), whereas the amount of RNA directly involved in protein synthesis, is known to vary with age, life-stage, organism size, disease-state and with changing environmental conditions (Bulow, 1970). The basic principle of using RNA/DNA ratio as a measure of health condition in an organism is that total RNA content is primarily a function of ribosome number and is positively correlated with new protein synthesis, whereas DNA content remains constant in an individual because it is a function of chromosome number (Buckley 1984; Buckley *et al.*, 1999; Dahlhoff, 2004; Chícharo & Chícharo 2008). Thus, organisms in good condition tend to have higher RNA: DNA ratios than do those in poor condition (Bulow, 1987). In fact RNA: DNA ratios have been used on a wide range of marine organisms, mainly phytoplankton (Dortch *et al.*, 1983), zooplankton (Sutcliffe, 1965; Ikeda *et al.*, 2007) and larval fish

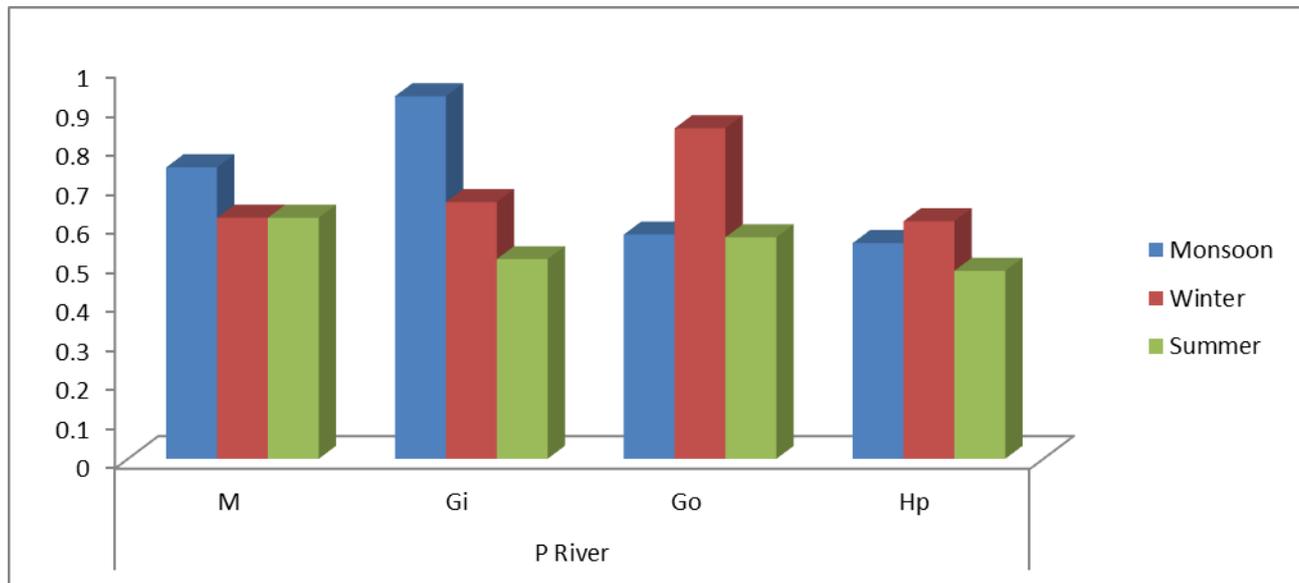
(Bulow, 1987; Buckley, L. 1984; Caldarone *et al.*, 2003) but also juvenile and adult fish (Bulow, 1970; Thorpe *et al.*, 1982), in bivalves (Chícharo *et al.*, 2001; Grémare, & Vétion, 1994; Chícharo & Chícharo, 1995) cephalopods (Clarke *et al.*, 1989; Sykes *et al.*, 2004) and crustaceans (Chícharo *et al.*, 2007; Lemos *et al.*, 2002) shown to reflect the nutritional status and may be useful in monitoring their physiological state in the field. An attempt has been made to understand seasonal changes in RNA: DNA ratio of gonad of male and female *L marginalis* during reproduction, inhabiting at Godavari River at Paithan been carried out.

MATERIALS AND METHODS

The Godavari is the second longest river in India. it flows through Paithan of Aurangabad district. Maharashtra At Paithan the river banks and Nath Sagar dam are sites for *Lamellidens marginalis*. Ten individuals of bivalve *Lamellidens*

marginalis from Godavari River and *Lamellidens corrianus* from Nandrabad pond (6.5 cm in shell length) were collected from collection sites in different seasons summer, monsoon and winter. Animals were immediately brought to the laboratory and acclimatized for 24 hrs for defecation. Animals were dissected and gonad smears were observed under microscope in order to find male and female individuals. Gonad weighed and homogenized in 750 μ l of trizol. The extraction of RNA and DNA was followed as per manufacturers' protocol. The RNA dissolved in 20 μ l of DEPC water. The solution was passed through several times in order to dissolve RNA 8mM NaOH aliquot containing. DNA was mixed with water to the final volume of 20 μ l & measure the absorbance RNA and DNA concentrations were read without further dilution on Nanodrop at 260nm & at 286nm at Paul Herbert Centre for DNA barcoding and biodiversity and expressed in ng/ μ l.

Fig1: RNA: DNA ratio of Female *Lamellidens marginalis* in different seasons



RESULTS AND DISCUSSION

RNA: DNA ratios of male and female *L marginalis* differed significantly in different tissues. Higher RNA: DNA ratio was found in Gill tissues in monsoon season in female and in mantle in male. In summer season high RNA: DNA ratio was found in mantle tissue of male and in winter it in

hepatopancreas of male. When compared with male and female tissues independently higher ratios are found in mantle of male in monsoon, in the gill tissues of female in monsoon. In winter female gonads also had shown higher values of ratio than male gonads and in male hepatopancreas than females.

Fig2: RNA: DNA ratio of Male *Lamellidens marginalis* in different seasons

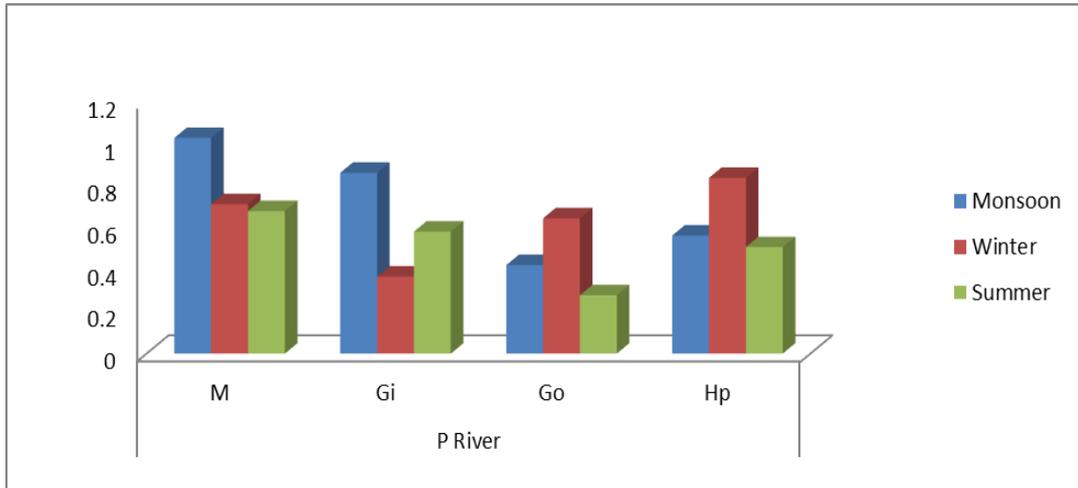


Fig3: RNA: DNA ratio of Male *Lamellidens corrianus* in different seasons

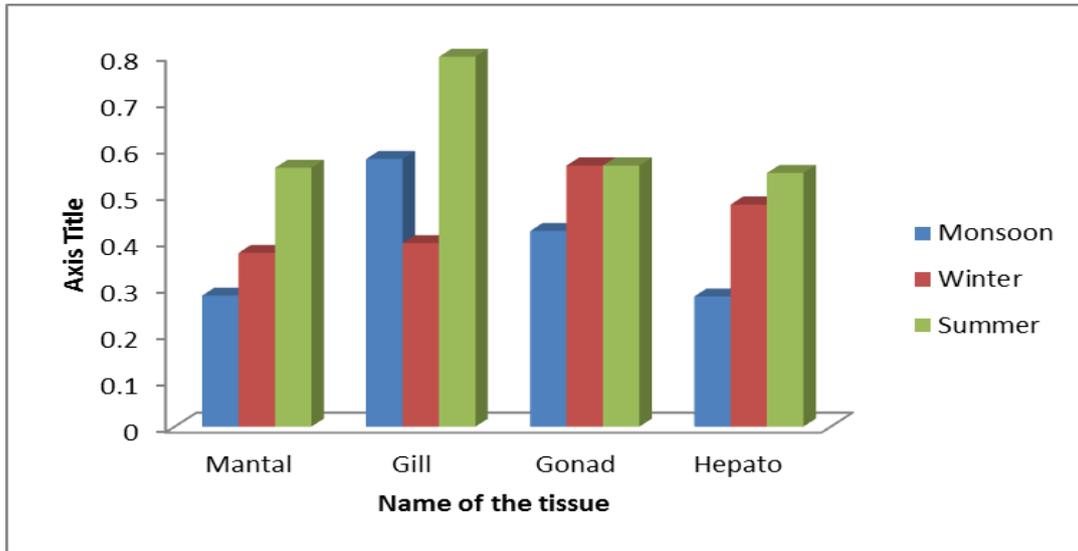
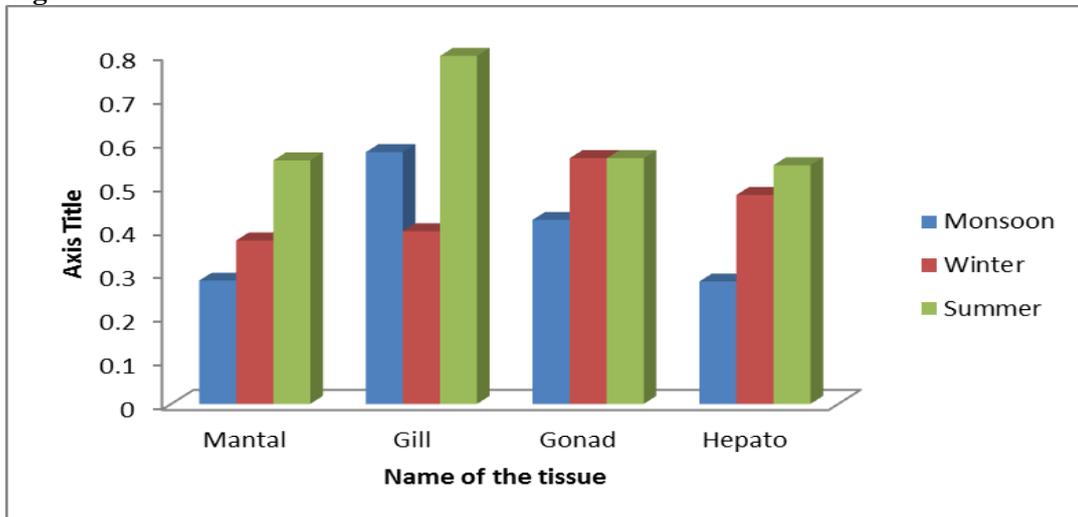


Fig4: RNA: DNA ratio of Female *Lamellidens corrianus* in different seasons



Much difference was observed in ratio in different tissues in the present study. This may be due to physiological changes occurring in the tissue according to the reproductive status of the animals. Changes in protein contents also seen in different tissues in different seasons maximum protein content was found in gonads in all seasons than in any other tissue. Mantle and hepatopancreas have shown nearly constant values, high protein content in mantle in winter (Jadhav and Jadhav, 2012).

RNA: DNA ratios of male and female *Lamellidens corrianus* differed significantly in different tissues. Higher RNA: DNA ratio is found in Gill and gonad tissues in monsoon season in female and in mantle if male. In summer season high RNA: DNA ratio is found in gill tissue of male and in winter it in hepatopancreas of male. When compared with male and female tissues independently higher ratios are found in gill of male in summer, in the all tissues of female in summer. In winter female gonads also had shown higher values of ratio than male gonads and in male hepatopancreas than females. Much difference is observed in ratio in different tissues in the present study. This may be due to physiological changes occurring in the tissue according to the reproductive status of the animals.

Maturation and reproduction, which differ between males and females, often require different amounts of energy, as discussed by Paon and Kenchington (1995) and Pérez *et al.* (2003). According to authors, reproductive costs are much greater for females than for a male, which implies that protein synthesis and therefore RNA content is greater in females than in males. The clam *R. decussates* (Pérez *et al.*, 2003) exhibits clear sexual differentiation in biochemical composition. Protein synthesis for females, especially during spawning activity, which could explain the higher RNA content and RNA: DNA ratios of female bivalves observed in the present study. However, this is not only restricted to spawning period: *C. crangon* was analyzed after spawning and in the resting period, and greater RNA content and RNA: DNA ratios were still evident in females. Behavior could also lead to differences in RNA: DNA ratios between sexes. According to Salgado *et al.*, (2004) higher RNA: DNA ratios of female *P.s microps* could be a consequence of differences in the activity patterns of sexes, because parental care is supported by males. Males are probably limited to foraging for food available around the nest during the breeding season, whereas females behave as food

maximizers. Feeding *R. decussate* exhibited the lowest RNA: DNA ratios, indicating very poor condition (Chicharo *et al.*, 2001), but the general pattern of low values for males and high values for females in terms of RNA concentrations and RNA: DNA ratios were still observed. In conclusion, caution is needed when interpreting RNA: DNA ratio data from adult organisms, because sex differences in nucleic acid concentrations bias RNA indices if the frequencies of sexes in samples are not representative of those in the population. Samples in which males are over-represented result in an underestimate of the condition of the population, and those in which females are over-represented result in an over estimate. Mitochondria and protein content are also greater in female rats than in males. Mean RNA: DNA ratios of females tended to be greater than those of males in all 3 species studied, largely owing to greater RNA concentrations Justo *et al.*, (2005). Robbins *et al.*, (1990), studied RNA: DNA ratio in male and female *P. maximus*, the authors reported that the RNA: DNA ratio of the male gonad was consistently ten times lower than that of the female gonad. In somatic tissues this could suggest a reduced protein synthetic activity. RNA: DNA ratios in spring and summer suggested that mussel physiology is h~ghlyplastic in response to site and area-related variations in environmental factors (Dahlhoff and Menge, 1996). Patterns of the RNA/DNA ratio are not always consistent across tissue types and food availability in different bivalve species (Mayrand *et al.*, 1994; Norkko & Thrush, 2006). Variation of RNA/DNA ratios of HP size effect was statistically significant in mussels only in which the RNA/DNA ratio decreased with increasing size. Small and medium mussels consistently showed the lowest RNA/DNA ratio when compared for two different sites (Yeung and Leung, 2013).

The seasonal variation in biochemical components is not related only to the reproductive cycle but is also influenced by the food availability and temperature as emphasized by Ansell (1974A, 1974B, 1974C, Darribaet. al., 2005). Seasonal changes in condition indices and biochemical components of the digestive gland, anterior adductor muscle, foot, and gonad of *Ensisarcuatus* (Jeffreys, 1865) were in relation to environmental conditions and reproductive events to meet the energy requirement. Gametes production occurs at the expense of other body reserves, from tissues like mantle and hepatopancreas when the gametes are

not ripe in *M. edulis* (Bayne and Thompson, 1970) and *P. viridis* (Nagabhushanam and Mane, 1978). Variations in organic constituents in different soft body parts of *L. marginalis* were observed by Sheikh (2011) in Pravara River in different seasons. In the present study the *L. marginalis* both the male and the female have shown beginning of gametogenic activity in the gonads during the summer season. The gonads showed accumulation of lipid globules and nutritive cells in the tissue. Proliferation of cells from germinal layer is also seen. Maturation of the gametes was observed during the monsoon season in both male and female gonads.

In the present study an attempt has been made to correlate the reproductive status of the bivalve molluscs and role of the other tissues in relation to RNA: DNA ratio. Since tissues of bivalves are contributing their role in the reproductive activities by accumulating and diverting biochemical substances as per need of gonad in different seasons. Some reports indicate mantle also plays important role in reproduction in movement of metabolites (Nagawanshi, 2013; 2014; Jadhav and Jadhav, 2012). Gill tissue plays an important role in housing the developing glochidia after fertilization by modifying it into marsupial (Nagawanshi, 1998). Thus the changes in RNA: DNA ratios in these tissues observed in different seasons can be correlated with the reproductive cycle of *L. marginalis*

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REFERENCES

Bayne BL and Thompson RJ, 1970: Some physiological consequences of keeping *Mytilusedulis* in the laboratory. *Helgol. Wiss. Meeresunter*, **20**: 526-552.
Bergeron JP, 1997. Nucleic acids in ichthyoplankton ecology: A review, with emphasis

on recent advances for new perspectives. *J. Fish Biol.*, **51**: 284-302.

Bergeron JP, 2000. Effect of strong winds on the nutritional condition of anchovy (*Engraulis encrasicolus* L.) larvae in the Bay of Biscay, Northeast Atlantic, as inferred from an early field application of the DNA/C index. – ICES. *Journal of Marine Science*, **57**: 249–255.

Buckley L, 1984. RNA: DNA ratio: an index of larval fish growth in the sea. *Mar. Biol.* **80**, 291-298.

Buckley LJ, Caldarone E and Ong TL, 1999. RNA-DNA ratio and other nucleic acid-based indicators for growth and condition of marine fishes. *Hydrobiol.* **401**: 265-277.

Bulow FJ, 1970. RNA-DNA ratios as indicators of recent growth rates of a fish. *J. Fish. Res. Board. Can.* **27**: 2343–2349.

Bulow JF, 1987. RNA-DNA ratios as indicators of growth rates in fish: A review. In *The age and growth of fish*; Summer felt. R.C. Hall G.E. Eds. The Iowa State University Press: Ames, Iowa, pp 45-64.

Caldarone EM, Buckley LJ, 1991: Quantitation of DNA and RNA in crude tissues extracts by flow injection analysis. *Anal. Biochem.* **199**: 137–141.

Canino MF, Caldarone EM, 1995: Modification and comparison of two fluorometric techniques for determining nucleic acid contents of fish larvae. *Fish. Bull.* **93**: 158–165.

Clarke A, Rodhouse PG, Holmes LJ, Pascoe PL, 1989: Growth rate and nucleic acid ratio in cultured cuttlefish, *Sepia officinalis* (Mollusca: Cephalopoda). *J. Exp. Mar. Biol. Ecol.* **133**: 229-240.

Clemmesen C, 1993. Improvements in the fluorometric determination of the RNA and DNA content in individual marine fish larvae. *Marine Ecology Progress Series* **100**: 177-183.

Chícharo L, Chícharo MA, 1995, The RNA: DNA ratio as useful indicator of the nutritional condition in juveniles of *Ruditapes decussatus*. *Sci. Mar.*, **59**: (Supl. 1) 95-101.

Chícharo L, Chícharo MA, Alves F, Amaral A, Pereira A, Regala J 2001. Diel variation of the RNA:DNA ratios in *Crassostrea angulata* (Lamarck) and *Ruditapes decussatus* (Linnaeus 1758) (Mollusca: Bivalvia). *J. Exp. Mar. Biol. Ecol.* **259**: 121–129

Chícharo, MA, 1996. Métodos de avaliação do estado nutricional em larvas de *Sardinapilchardus* (Walbaum, 1792) aplicados ao estudo das condições de sobrevivência no meio natural.

Unpublished Ph.D. thesis, University of Algarve, Faro.

Chícharo, MA, Chícharo L, Amaral A, Morais P, 2007. Sex effect on ratios and concentrations of DNA and RNA three in marine organisms. *Mar. Ecol. Prog. Ser.* **332**: 241-245.

Dahlhoff EP, B A Menge, 1996. Influence of phytoplankton concentration and wave exposure on the ecophysiology of *Mytilus californianus* us. *Mar. Ecol. Prog. Ser.* **144**:97-107.

Dortch Q, Roberts TL, Clayton JR Jr, Ahmed SI, 1983. RNA/DNA ratios and DNA concentrations as indicators of growth rate and biomass in planktonic organisms. *Mar. Ecol. Prog. Ser.* **13**: 61-71.

Grapputo A, S Boman, L Lindstrom, A Lyytinen, J Mappes, 2005. The voyage of an invasive species across continents: genetic diversity of North American and European Colorado potato beetle populations. *Molecular Ecology*, **14** (14): 4207-4219.

Grémare A, Ve' tion G, 1994. Comparison of several spectrofluorimetric methods for measuring RNA and DNA concentrations in the deposit-feeding bivalve *Abra ovata*. *Comp. Biochem. Physiol.* **107**(B): 297-308.

Houlihan DF, Hall SJ, Gray C, Noble BS, 1988. Growth rates and protein turnover in Atlantic cod, *Gadus morhua*. *Can. J. Fish Aquat. Sci.*, **45**: 951-964.

Ikeda T, San F, Yamaguchi A, Matsuishi T, 2007. RNA: DNA ratios of calanoid copepods from the epipelagic through abyssopelagic zones of the North Pacific Ocean. *Aquatic Biol.*, **1**: 99-108.

Jadhav MR and BN Jadhav, 2012. Changes in the protein content of *Lamellidens marginalis* from Jayakwadi dam at paithan during different seasons (M.S) India. *International Multidisciplinary Research Journal*, **2**(10):01-02

Justo R, M Frontera, E Pujol, S Rodríguez-Cuenca, I Lladó, F José García-Palmer, P Roca, M. Gianotti, 2005. Gender-related differences in morphology and thermogenic capacity of brown adipose tissue mitochondrial subpopulations. *Life Sciences*, **76** (10): 1147-1158.

Kelley N R, Ashwood-Smith M J and Ellis DV, 1982. Duration and timing of spermatogenesis in a stock of the mussel *Mytilus californianus*. *J. Mar. Biol. Ass. U.K.* **62**: 509-519.

Lemos D, Garcia-Carren FL, Hernandez P, Toro AN 2002. Ontogenetic variation in digestive proteinase activity, RNA and DNA content of larval

and postlarval white shrimp *Litopenaeus schmitti*. *Aquaculture*, **214**: 363-380.

Lucas A, and Beninger P G, 1985. The use of physiological condition indices in marine bivalve aquaculture, *Aquaculture*, **44** (3): 187-200.

Mayrand E, Pellerin-Massicotte J and Vincent B, 1994. Small scale variation of biochemical indices of growth in *Mya arenaria* (L.). *Journal of Shellfish Research*, **13**: 199-205.

McGurk MD, Warburton HD, Galbraith M, Kusser WC, 1992. RNA/ DNA ratio of herring and sand lance larvae from Port Moller, Alaska. Comparison with prey concentration and temperature *Fish. Oceanogr.* **1** (3):193-207.

Nagawanshi M, 1998. *Reproductive physiology of freshwater bivalve molluscs from Aurangabad*, Maharashtra State Ph. D. Thesis. Dr. Babasaheb Ambedkar Marathwada University, Aurangabad.

Nagawanshi M, 2013. Seasonal variations in the nucleic acids and RNA:DNA ratio in the gonad of the freshwater bivalve *L. corrianus* from Nandrabad pond, *Trends In Life Sciences*, **2** (4):36-38.

Nagawanshi M, 2014. Seasonal variations in the nucleic acid and RNA:DNA ratio in the gonad of the freshwater bivalve *l. marginalis* from Godavari river. *National Journal of Life Sciences*, **11**(1):53-55.

Nagabhushanam R and U H Mane, 1978. Seasonal variation in the biochemical composition of *Mytilus viridis* at Ratnagiri on the West Coast of India. *Hydrobiologia*, **57**(1): 69-72.

Norkko J and Thrush SF, 2006. Ecophysiology in environmental impact assessment: implications of spatial differences in seasonal variability of bivalve condition. *Marine Ecology Progress Series*, **326**:176-186.

Paon L A and Kenchington E L R. 1995. Changes in somatic and reproductive tissues during artificial conditioning of the sea scallop, *Placopecten magellanicus* (Gmelin, 1791). *J. Shellfish. Res.* **14**:53-58.

Pérez Camacho A, Delgado M, Fernández-Reiriz MJ and Labarta U, 2003. Energy balance, gonad development and biochemical composition in the clam *Ruditapes decussatus*. *Mar. Ecol. Prog. Ser.* **258**:133-145.

Robbins L, P Lubet and JY Besnard, 1990. Seasonal variations in the nucleic acid content and RNA:DNA ratio of the gonad of the scallop *Pecten maximus*. *Marine Biol.* **105**: 191-195.

Regnault M, Luquet P 1974. Study by evolution of nucleic acid content of prepubertal growth in the shrimp *Crangon vulgaris*. *Mar. Biol.* **25**:291-298.

- Salgado JP, Cabral HN, Costa MJ, 2004**, Feeding ecology of *Pomato schistus minutus* (Pallas, 1770) and *Pomato schistus microps* (Krøyer, 1838) in the upper Tagus estuary. *Sci. Mar.* **68**:425–434.
- Sambrook J and Russel DW, 2001**. Molecular Cloning, A Laboratory Manual Vol. 1, 2, 3, Cold Spring Harbor Laboratory Press, New York
- Sokolov E P, 2000**. An improved method for DNA isolation from mucopolysaccharide-rich molluscan tissue. *Journal of Molluscan Studies*, **66**: 573-575.
- Sutcliffe WH Jr., 1965**. Growth estimates from ribonucleic acid content in some small organisms. *Limnol. Oceanogr.* **10**: 253-258.
- Sykes A, Domingues P, Andrade JP. 2004**. Nucleic acid derived indices or instantaneous growth rate as tools to determine different nutritional condition in cuttlefish (*Sepia officinalis*, Linnaeus 1758) hatchlings. *J. Shell. Res.*, **23**: 585-591.
- Thorpe JE, Talbot C, Villarreal, C. 1982**. Bimodality of growth and smolting in Atlantic salmon, *Salmo salar L.* *Aquaculture*, **28**: 123-132.
- Timmermans MJ, J Ellers, J Marien, SC Verhoef, E B, Ferwerda, and N M Van Straalen, (2005)** Genetic structure in *Orchesellacincta* (Collembola): strong subdivision of European populations inferred from mtDNA and AFLP markers. *Molecular Ecology*, **14** (7): 2017-2024.
- Yeung J WY and Leung KMY, 2013**. Effects of animal size and nutritional status on the RNA/DNA ratio in different tissues of the green-lipped mussel *Perna viridis* *Journal of the Marine Biological Association of the United Kingdom*, **93**(1), 217–225.
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